

New MALDI-TOF MS with very high  
resolving power and mass accuracy

Presented at ASMS  
June 3, 2008

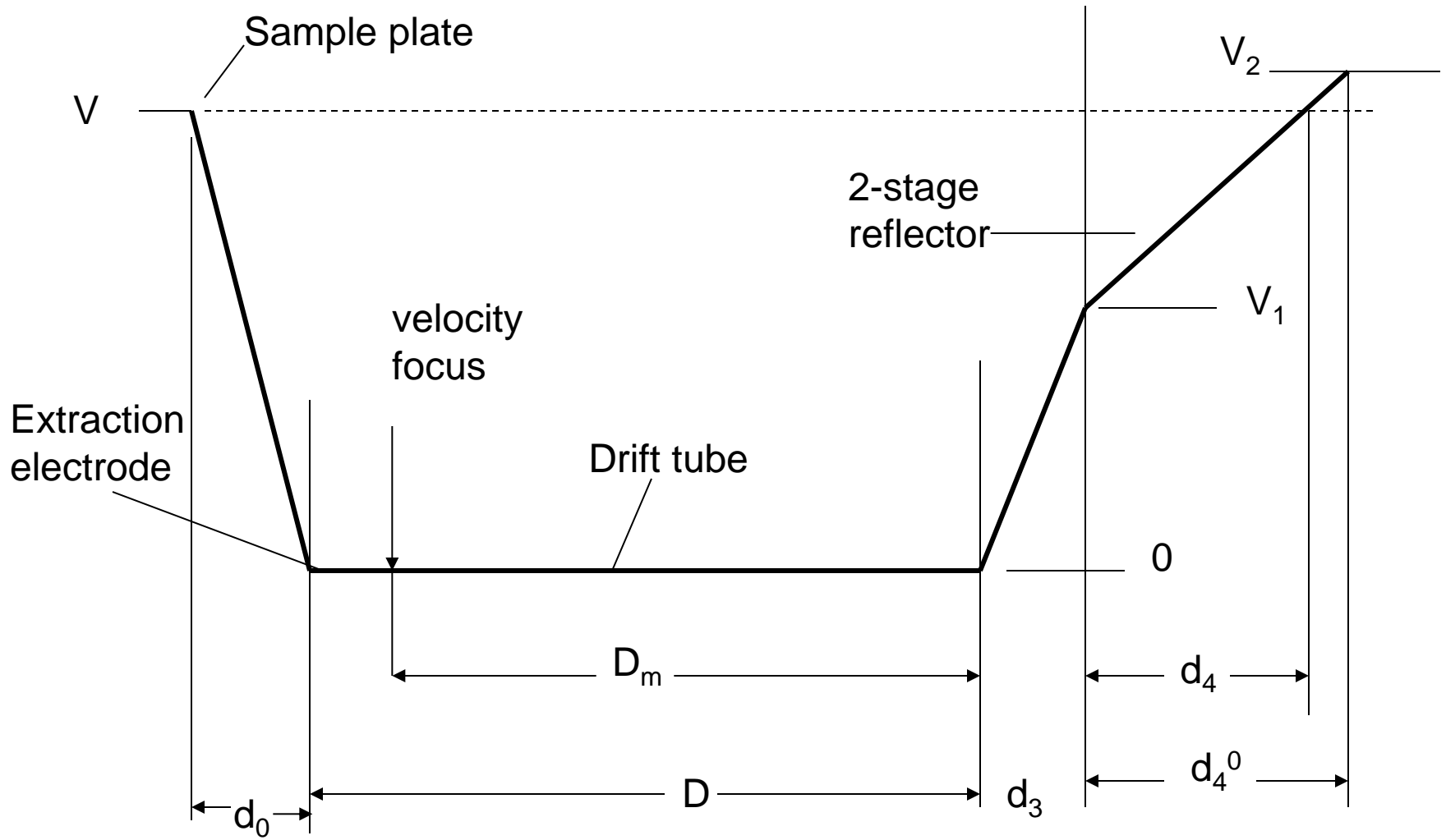
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# Objectives

- Determine optimum design parameters for a high-performance bench-top MALDI-TOF
  - Resolving power and mass accuracy
  - Sensitivity and dynamic range
  - High throughput
  - Efficient sample utilization
  - Simple and reliable
  - Low cost
- Build prototype instrument and compare experimental results with theoretical predictions

# Common Features of Virgin MALDI TOF

- Simplified sample loading with super microplate format (102x108 mm)
- Benchttop cabinet
  - 1.75m high x 0.5 m wide x 0.7 m deep
  - 100 kg max weight
- 5 kHz laser, up to 100 spectra/sec
- Fully automated, no operator expertise required
  - No joystick, no camera
- 24/7 operation with minimal preventative maintenance



Potential diagram for high-resolution TOF with single-field source and two-field reflector. The field-free distance  $D$  is the distance from the source to the mirror plus the distance from the mirror to the detector (not shown).

## **Instrument Parameters for Reflecting Analyzer (single-field source and two-field mirror)**

$D_e=3200$ ,  $d_0=6$ ,  $D=2264$ ,  $d_3=114.2$ ,  $d_4^0=127.7$  all in mm

$V=8.56$ ,  $V_1=6.31$ ,  $V_2=9.23$  all in kV

$\delta t=1.5$  nsec (0.5 nsec bins, 1 nsec single ion pulse width)

Focus mass  $m^*=3$  kDa

$D_v=6d_0=36$  mm,  $v_n$  (for  $m^*$ )=0.0235 mm/nsec

Time lag  $\Delta t=d_0/v_n=250$  nsec

$K=2d_0/(D_v-D_s)=0.5$

## **Initial Conditions for MALDI (typical)**

$\delta v_0=400$  m/sec,  $\delta x=0.01$  mm

**Trajectory error and voltage error assumed to be small**

# References

- M. L. Vestal and P. Juhasz, “Resolution and Mass Accuracy in Matrix-Assisted Laser Desorption Time-of-Flight Mass Spectrometry”, *J. Am. Soc. Mass Spectrom.* **9**, 892-911 (1998).
- M. L. Vestal and K. Hayden, “High-performance MALDI-TOF mass spectrometry for proteomics”, *Int. J. Mass Spectrometry* **268**, 83-92 (2007).
- M. L. Vestal and K. Hayden, “High-performance MALDI-TOF Mass Spectrometry”, paper TOFam08:15 presented at 55th ASMS Conference on Mass Spectrometry and Allied Topics, Indianapolis, June 3-7, 2007.
- S. J. Hattan and M. L. Vestal, “Novel 3-D sample plate using monolithic capture media in collimated-hole structures for interfacing high capacity separations with MALDI-TOF”, poster TBP-060 presented at 56th ASMS Conference on Mass Spectrometry and Allied Topics, Indianapolis, June 3, 2008.
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*Contributions to relative peak width,  $\Delta m/m$ , with 1<sup>st</sup> and 2<sup>nd</sup> order velocity focusing*

Initial position,  $\delta x$ :  $R_{s1} = 2[(D_v - D_s)/2d_0y](\delta x/D_e)$

Initial velocity,  $\delta v_0$ :  $R_{v1} = (4d_0y/D_e)(\delta v_0/v_n)[(1 - (m/m^*)^{1/2})]$

$R_{v2} = 2[2d_0y/(D_v - D_s)]^2 (\delta v_0/v_n)^2 = 0$

$R_{v3} = 2[2d_0y/(D_v - D_s)]^3 (\delta v_0/v_n)^3$

$R_v = R_{v1} + R_{v3}$

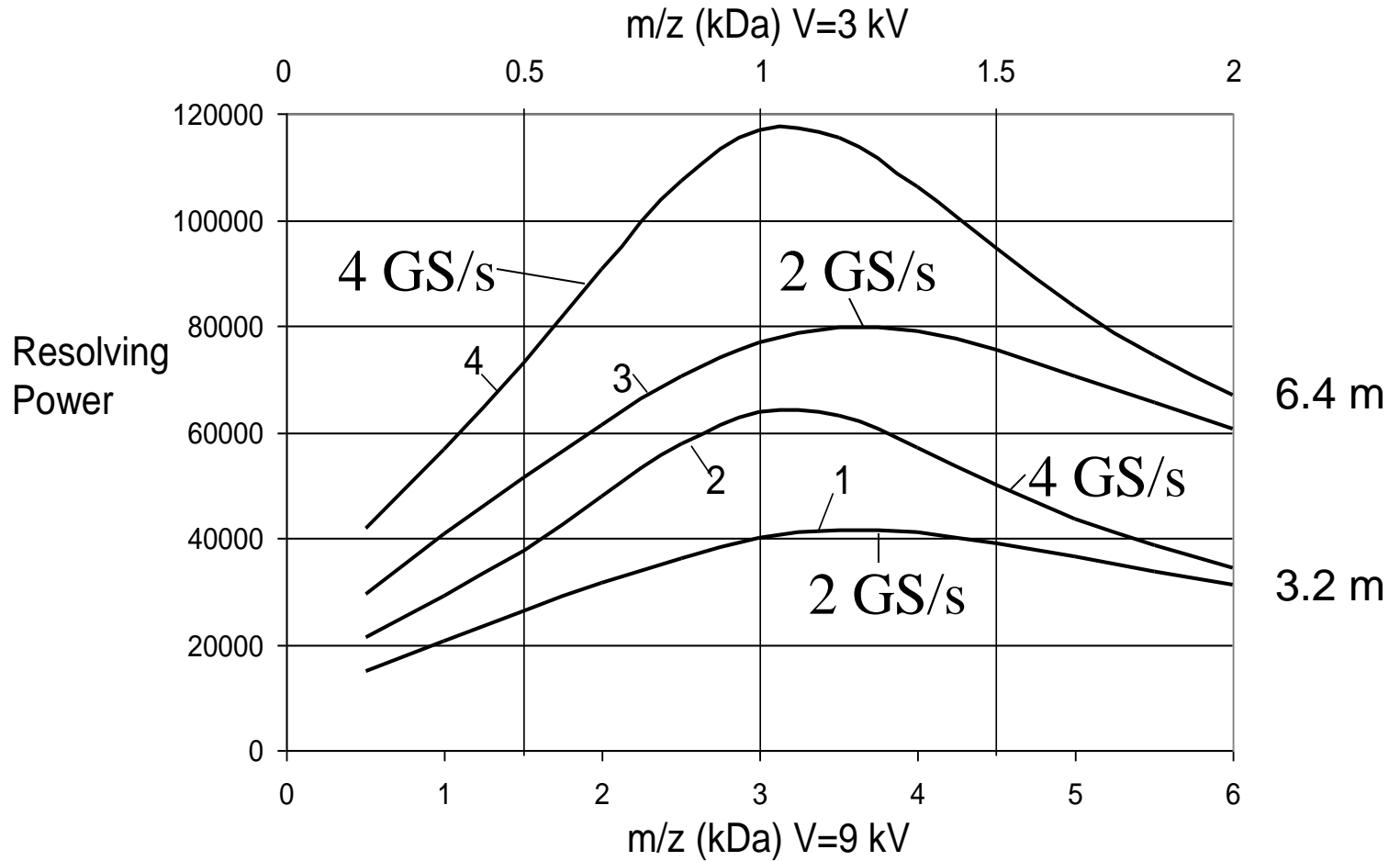
Time error,  $\delta t$ :  $R_t = 2\delta t/t = 2\delta t v_n/D_e$

Trajectory error,  $\delta L$ :  $R_L = 2\delta L/D_e$  } Neglected in initial calculation

Voltage error,  $\delta V$ :  $R_V = \delta V/V$  } calculation

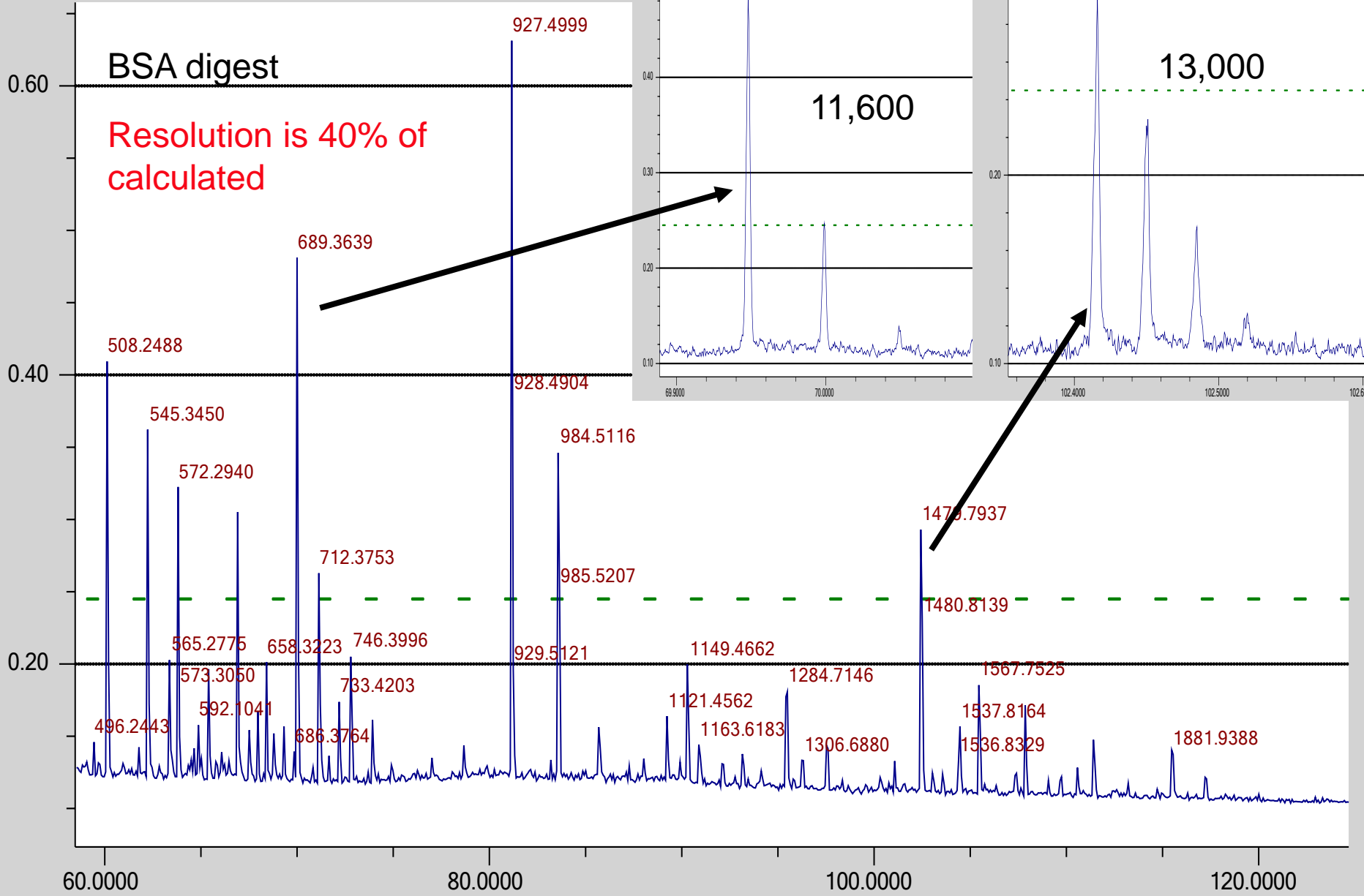
Resolving power:

$$R^{-1} = [R_{s1}^2 + R_v^2 + R_t^2 + R_L^2 + R_V^2]^{-1/2}$$

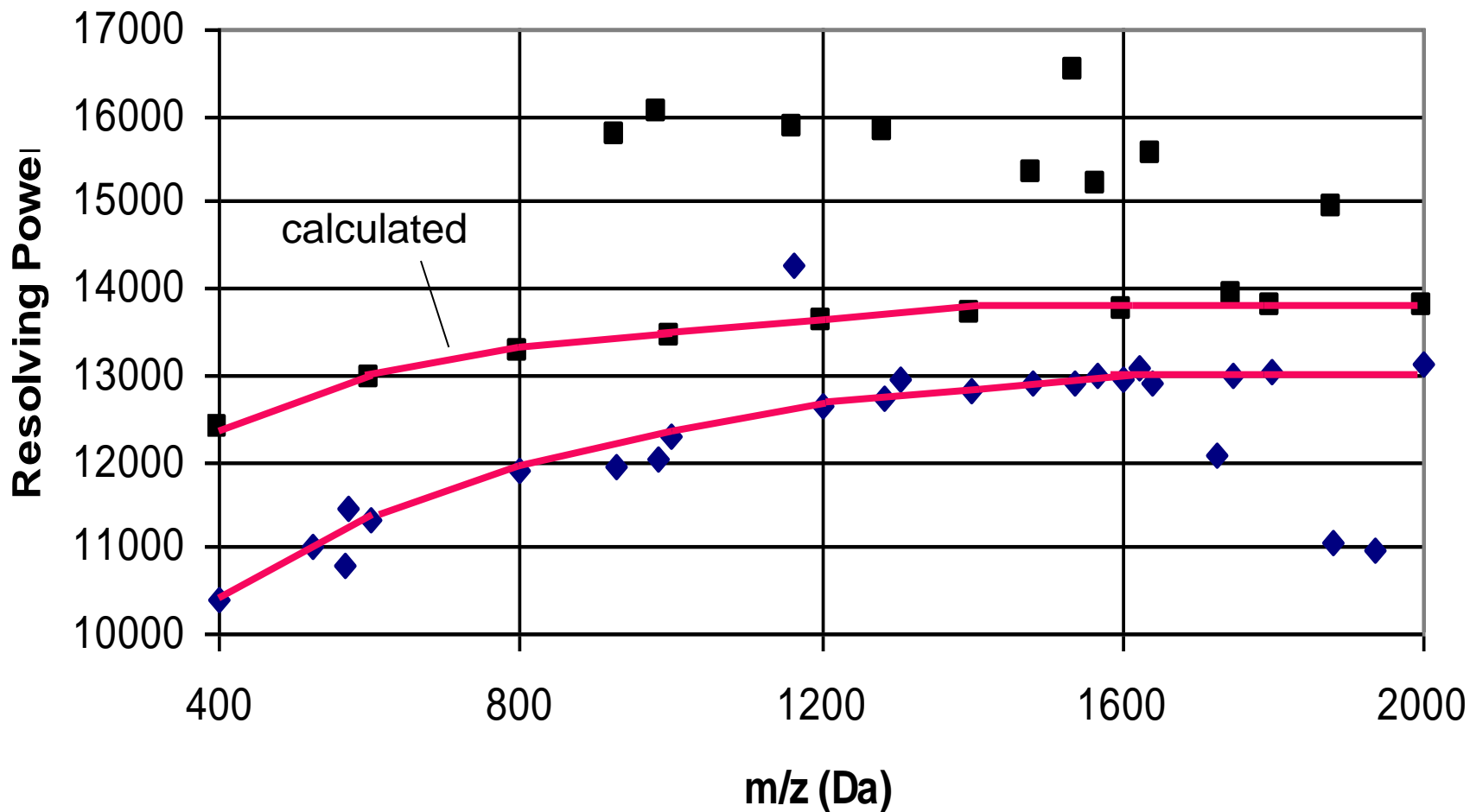


Calculated resolving power as function of m/z, curve 1 corresponds to experimental instrument





Spectrum from original version



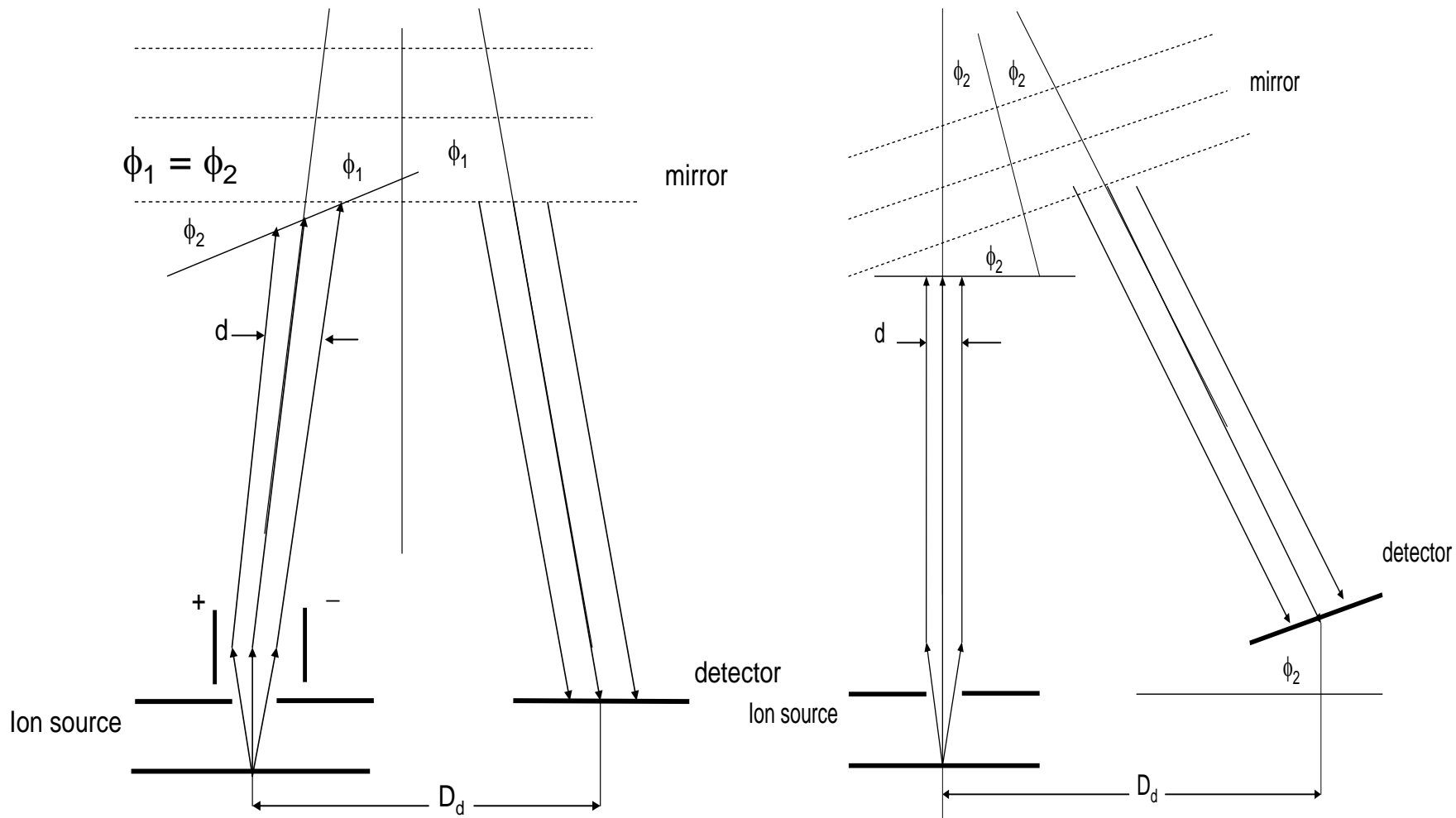
Comparison of calculated resolving power with experimental results  
 With  $\delta m/m=70$  ppm due to mass independent errors.

Measured  $\blacklozenge$  5  $\mu$ channel plate  $\blacksquare$  ETP MagnetTOF™ (DM167)

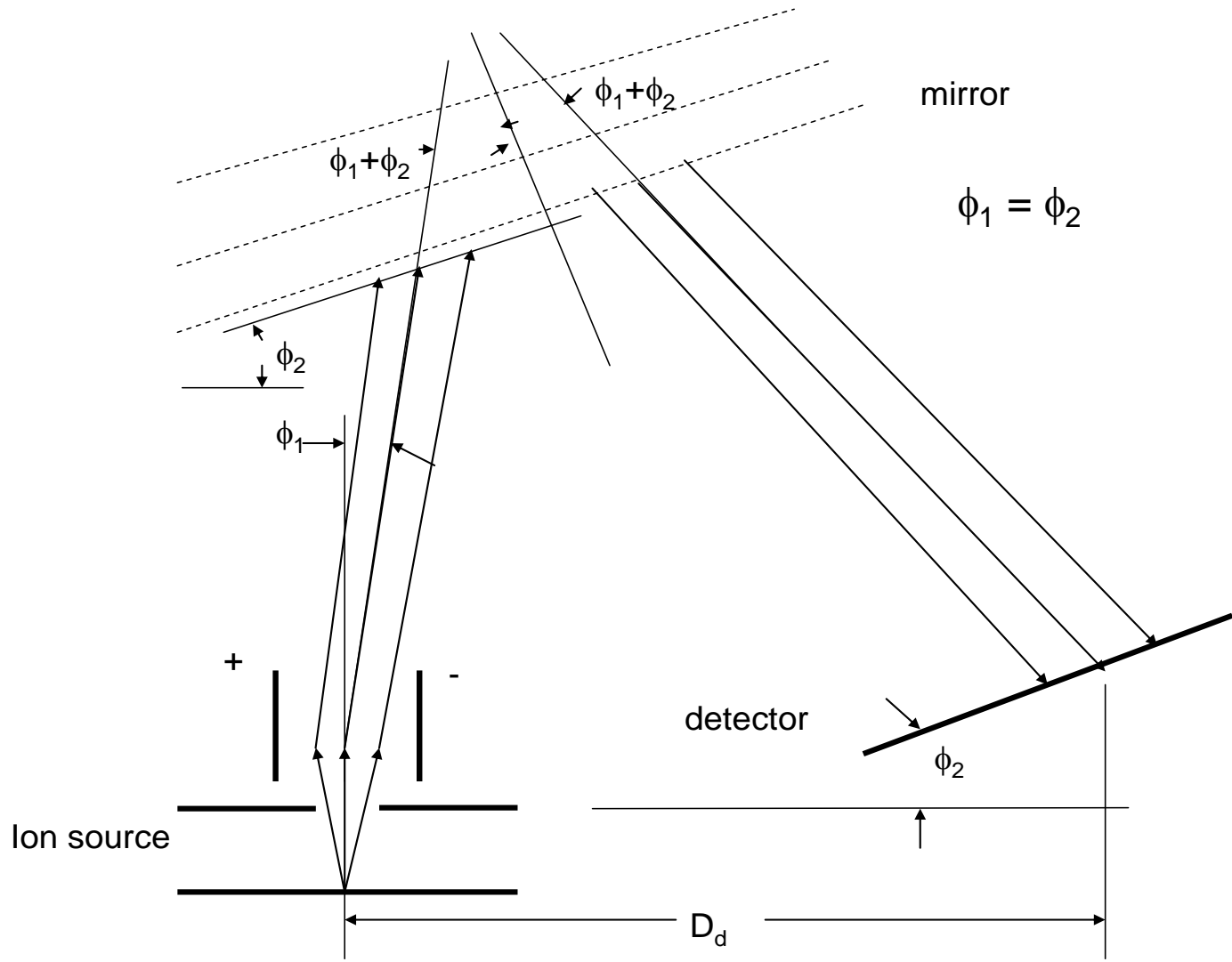
Original version before trajectory error correction

# Changes to correct mass independent errors

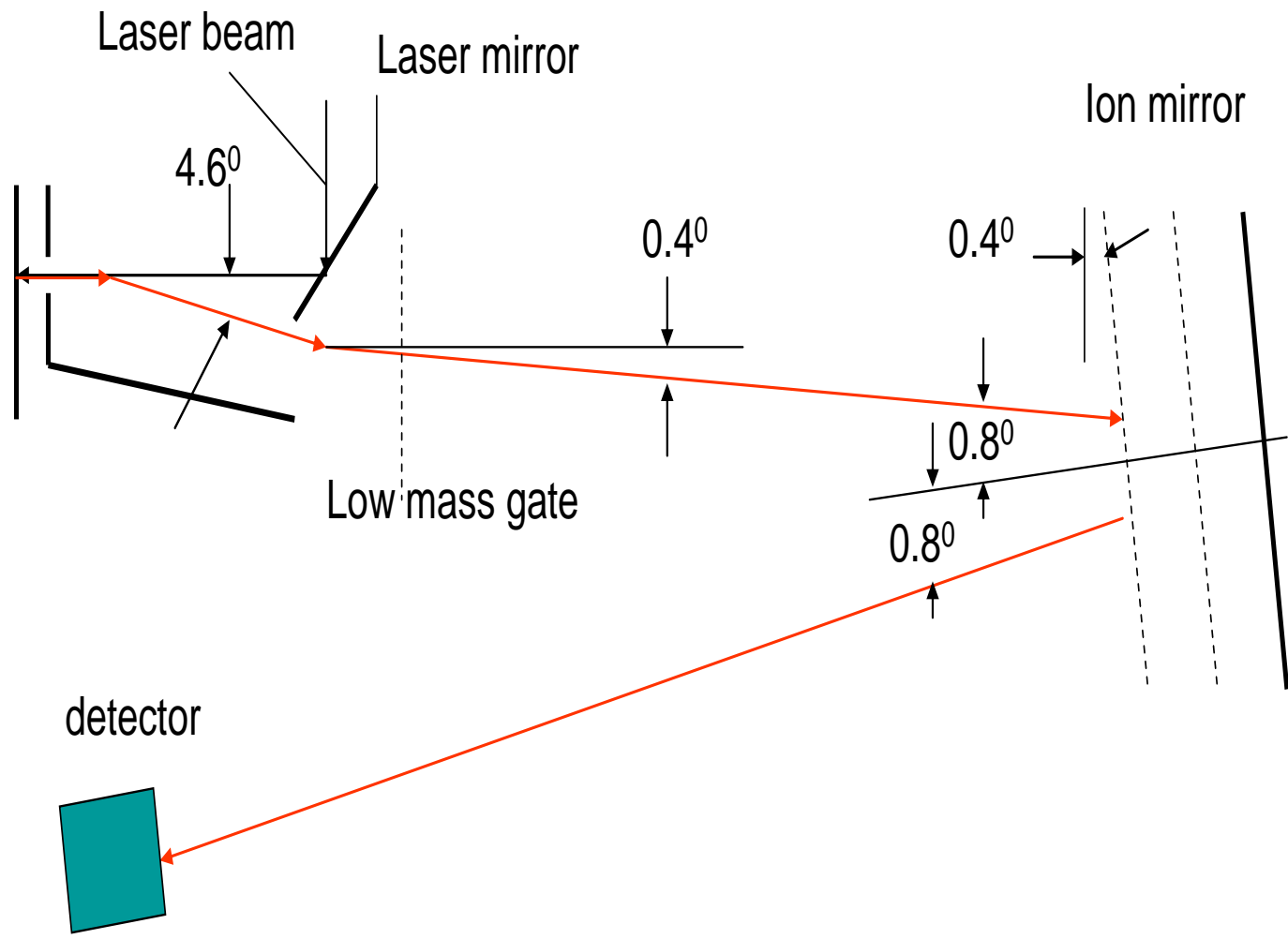
- Add voltage regulator at input to mirror HV supplies (removes low frequency noise)
- Correct trajectory error due to ion deflector
- Use faster detector (0.5 ns)
  - ETP DM167 in place of 5  $\mu\text{m}$  dual channel plate
- Use faster digitizer
  - 0.5 ns bins > 0.25 ns bins



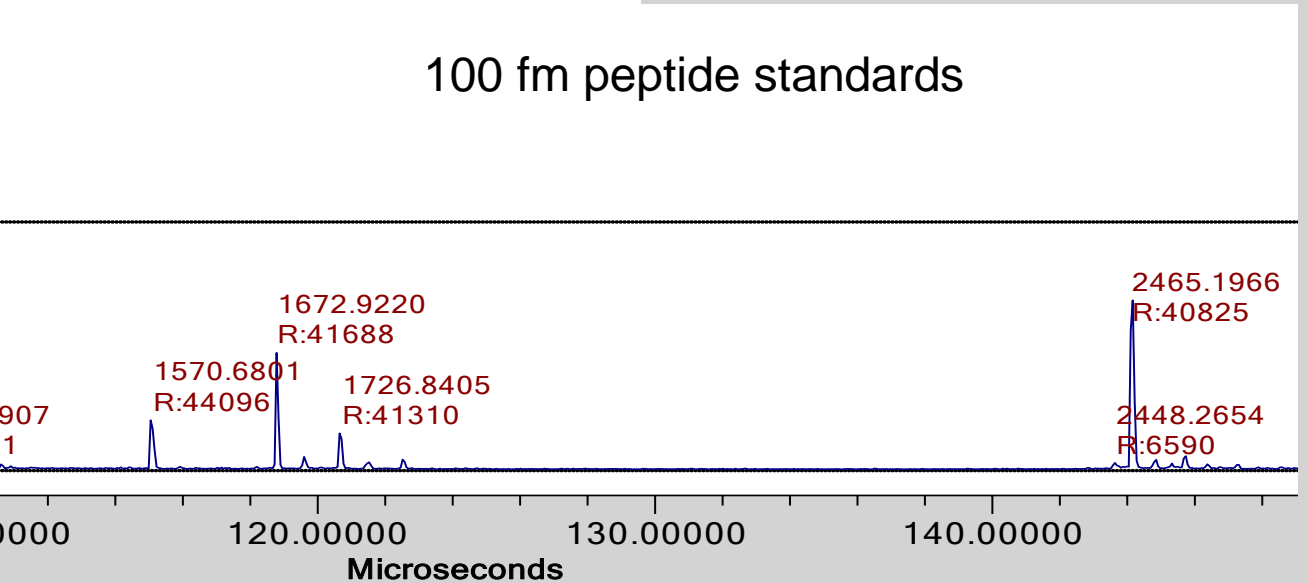
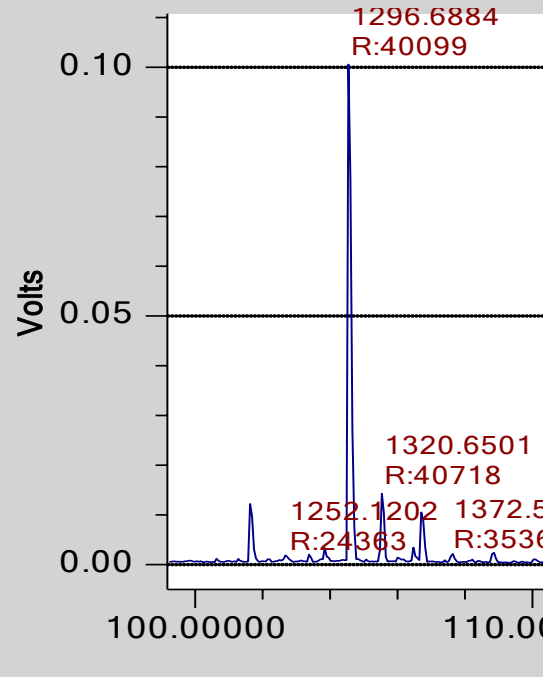
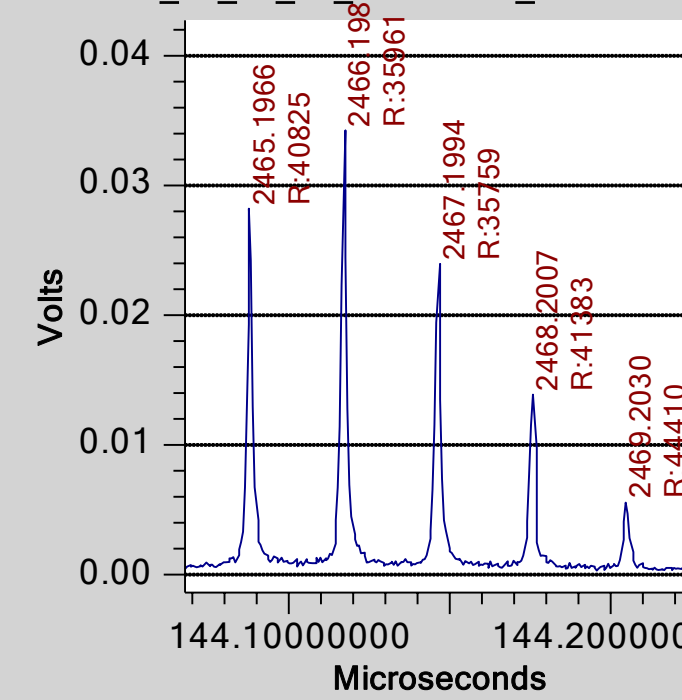
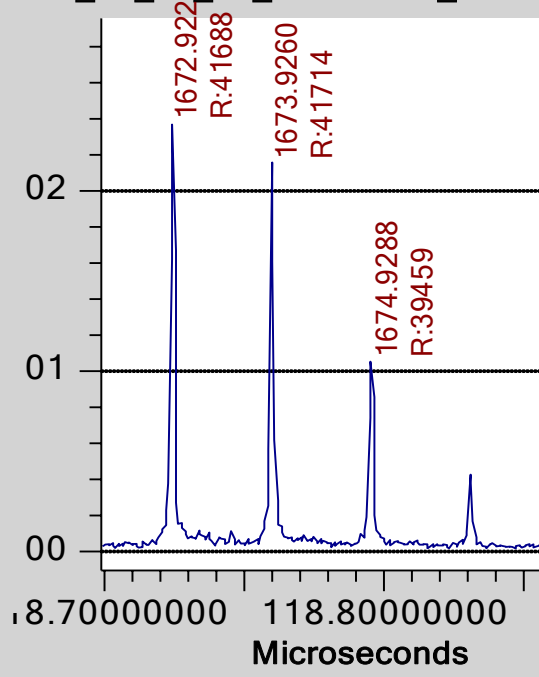
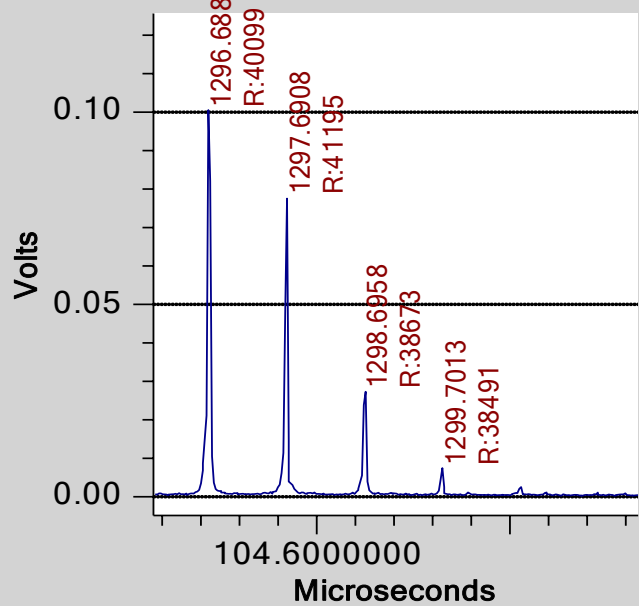
Ray diagrams indicating source of trajectory error



Ray diagram showing general correction to trajectory error due to deflection

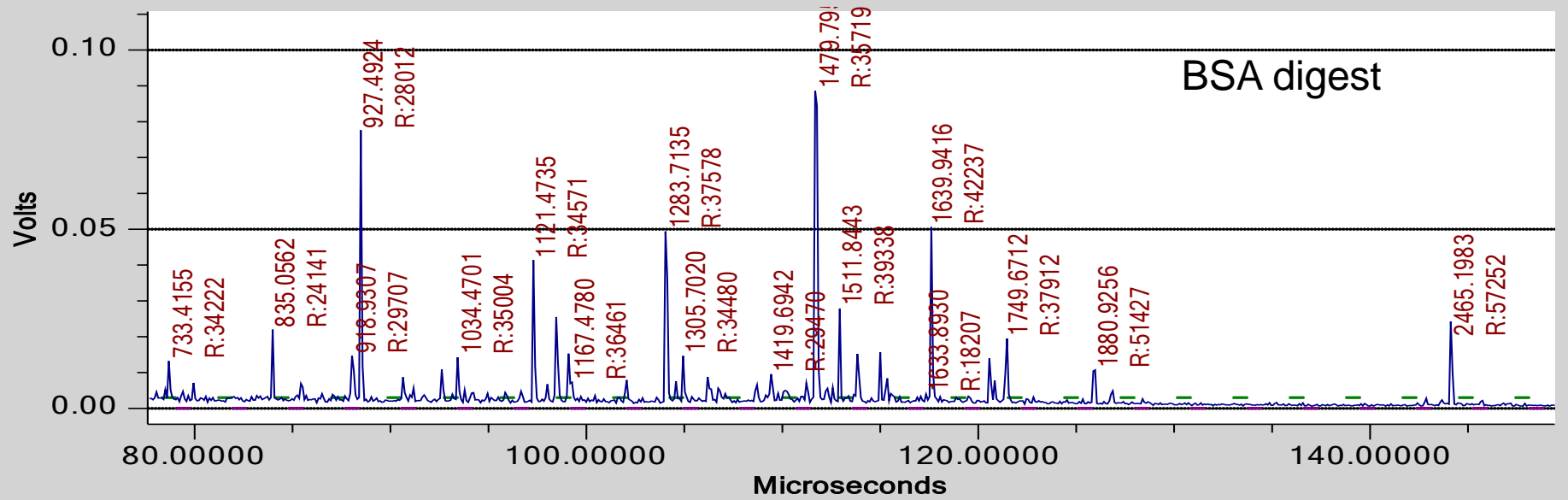
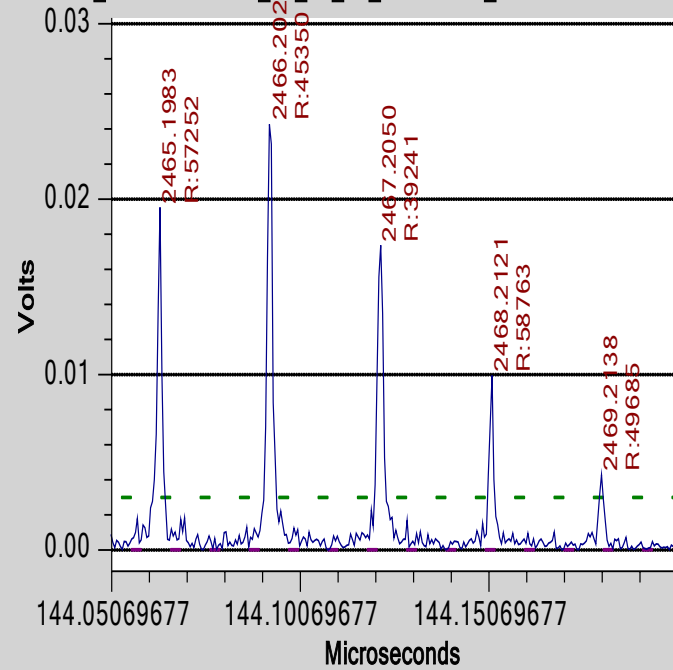
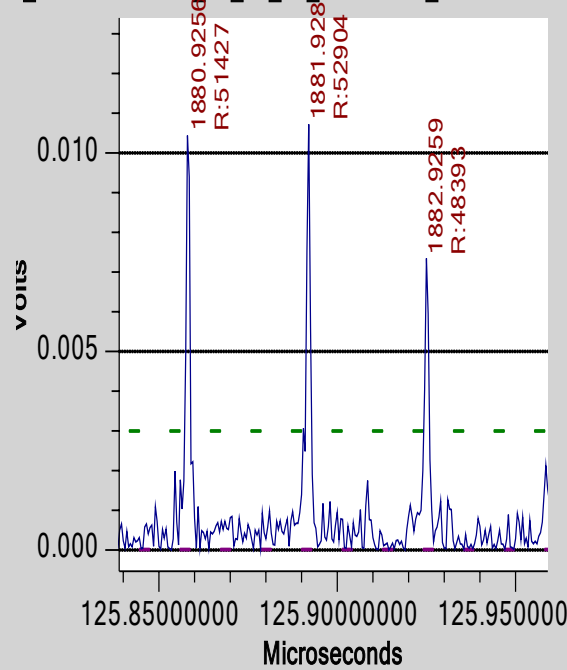
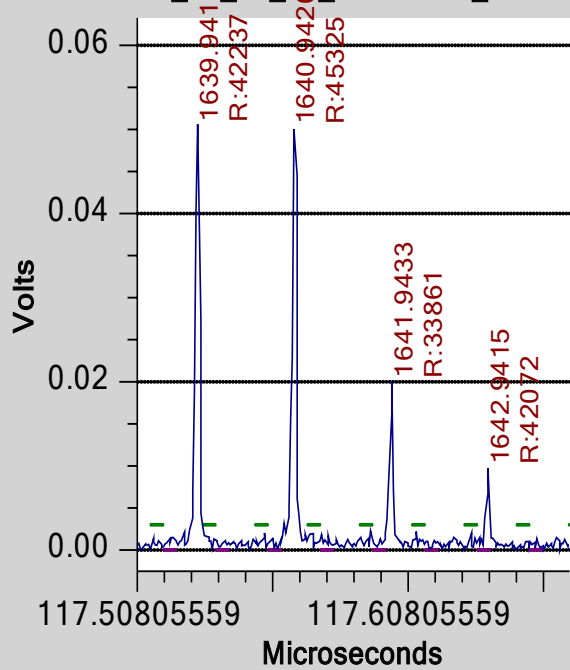


Ray diagram of final version employed in the corrected version of the experimental analyzer



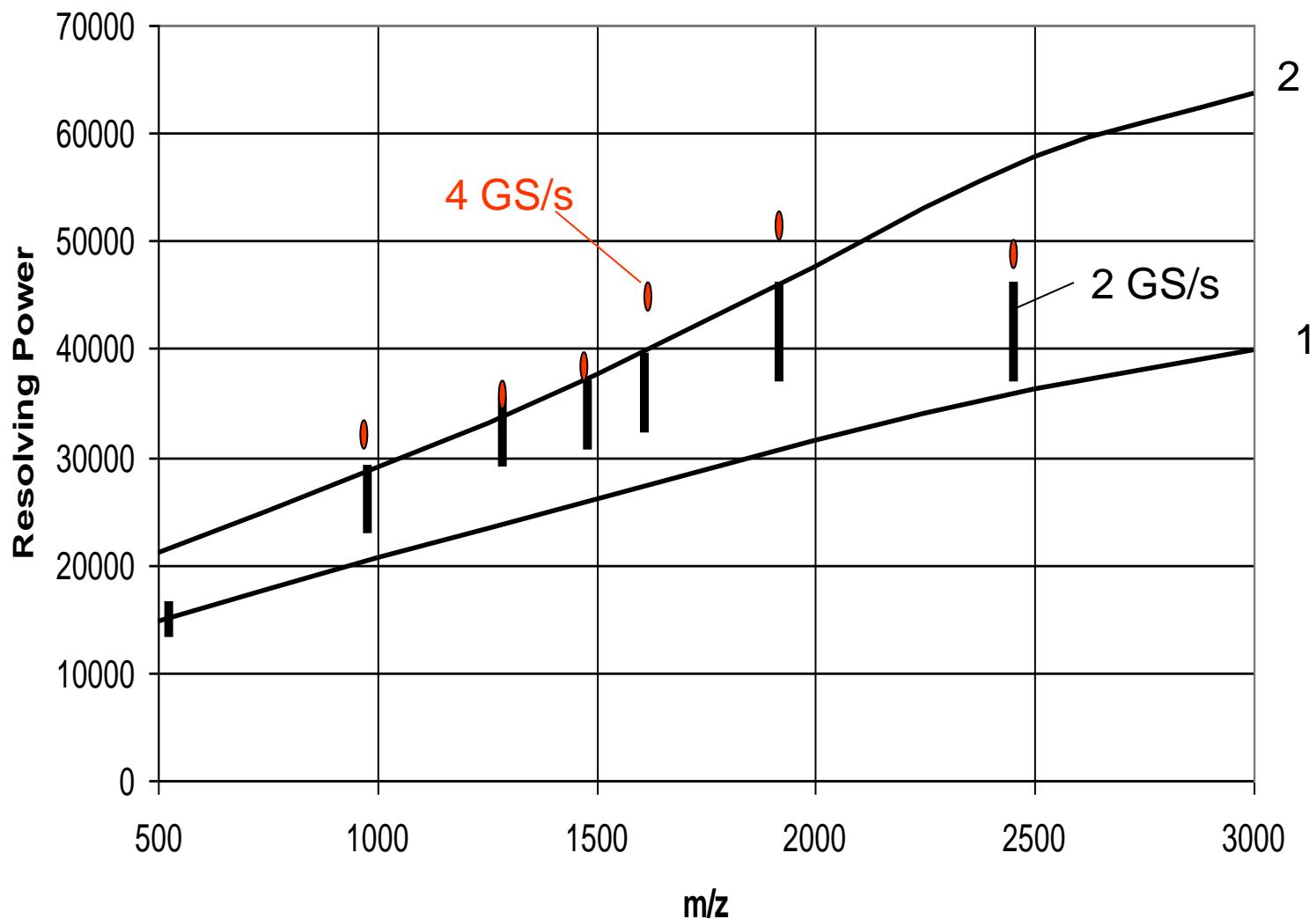
100 fm peptide standards

Example of spectrum obtained with corrected analyzer



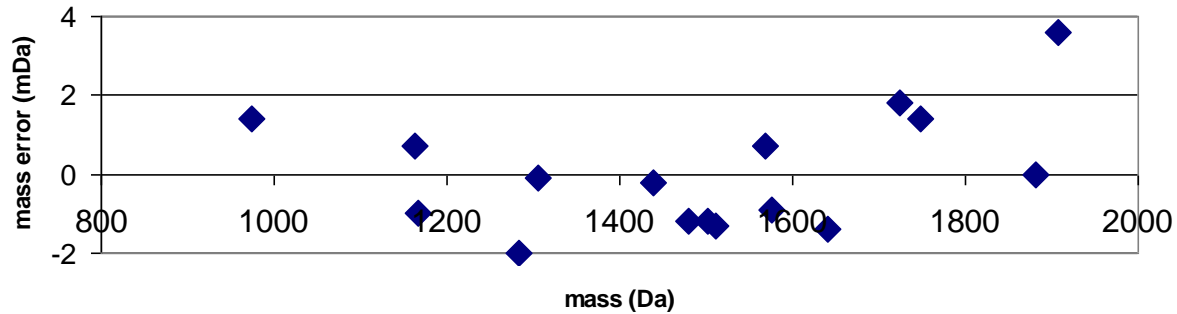
Example of spectrum obtained with corrected analyzer





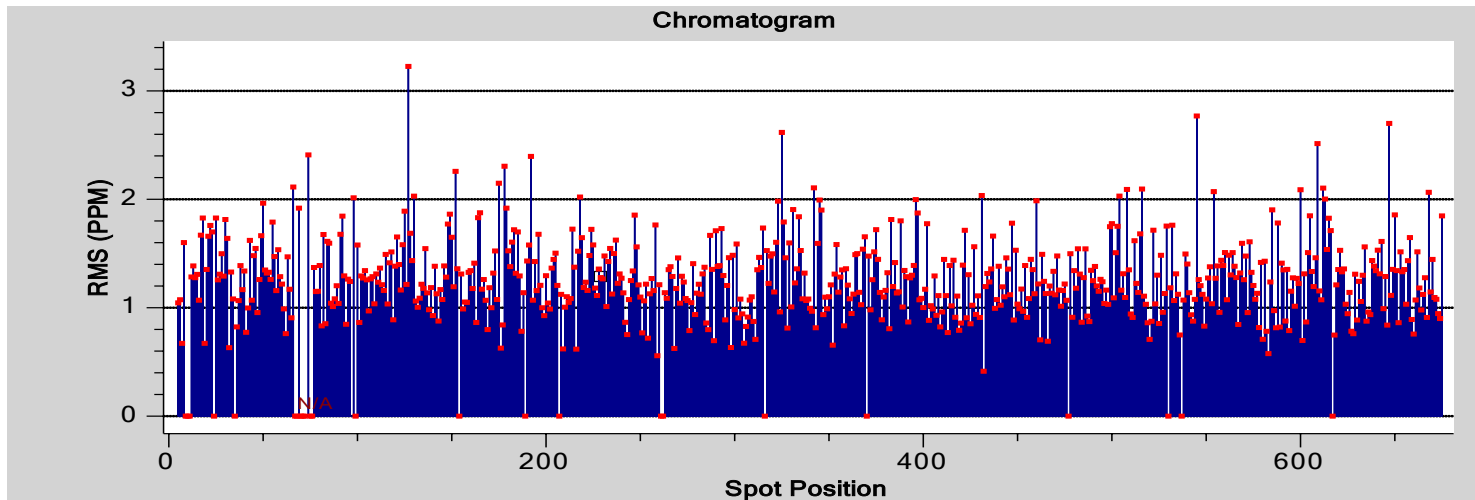
Comparison of experimental results with theoretical calculations. Bars are measured resolving power with standard deviation of 5 replicates. Curve 1 corresponds to digitizer with 0.5 ns bins and curve 2 to 0.25 ns bins.

Calibration Equation:  $m^{1/2} = D_0 + D_1 t [1 + D_2 t + D_3 t^2]$   
Determine coefficients by least square fit to multiple  
Peaks covering broad mass range



Average = -0.1 mDa

RMS = 0.99 ppm



RMS error for 10 peaks in spectrum of tryptic digest of BSA for all 675 spots on a 102x108 mm sample plate with automatic 2-point internal Calibration ( $D_0$  and  $D_1$ ) with  $D_3$  and  $D_4$  fixed.

# Serial Dilution Experiment

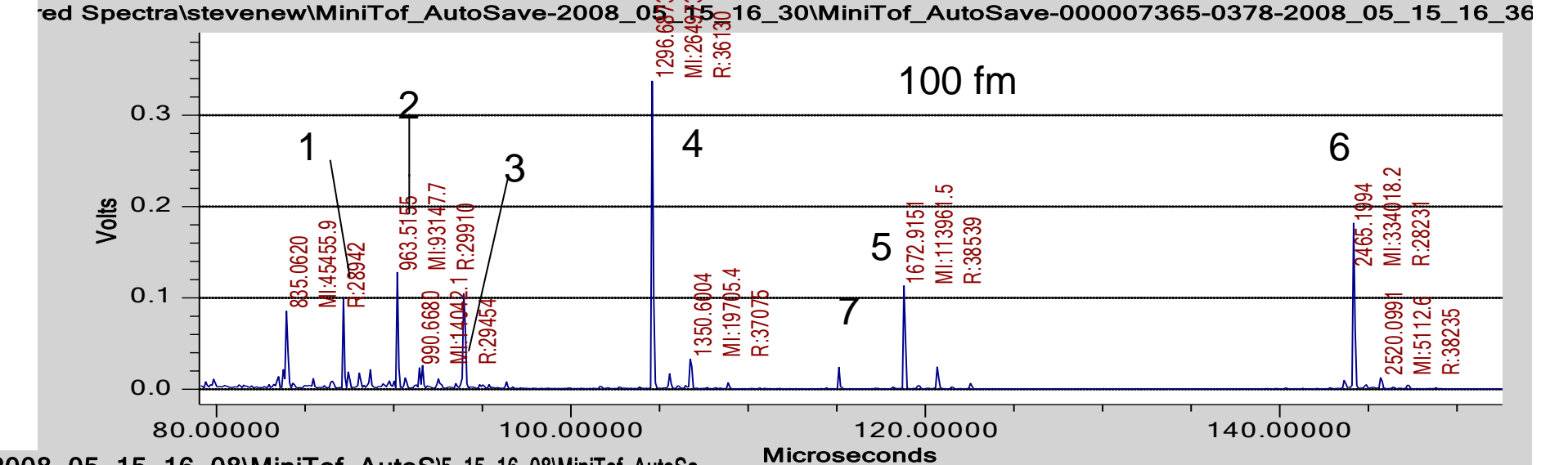
## Peptide Standards

	<b>MH<sup>+</sup></b>	<b>No.</b>
Angiotensin Fragment 1-7	899.47	1
des-Pro2 Bradykinin	963.51	2
Angiotensin II Acetate	1046.54	3
Angiotensin I	1296.69	4
Neurotensin	1672.92	5
ACTH (18-39)	2465.20	6
<b>Glu-1 Fibrinopeptide B</b>	<b>1570.68</b>	<b>7(&lt;10% pure)</b>

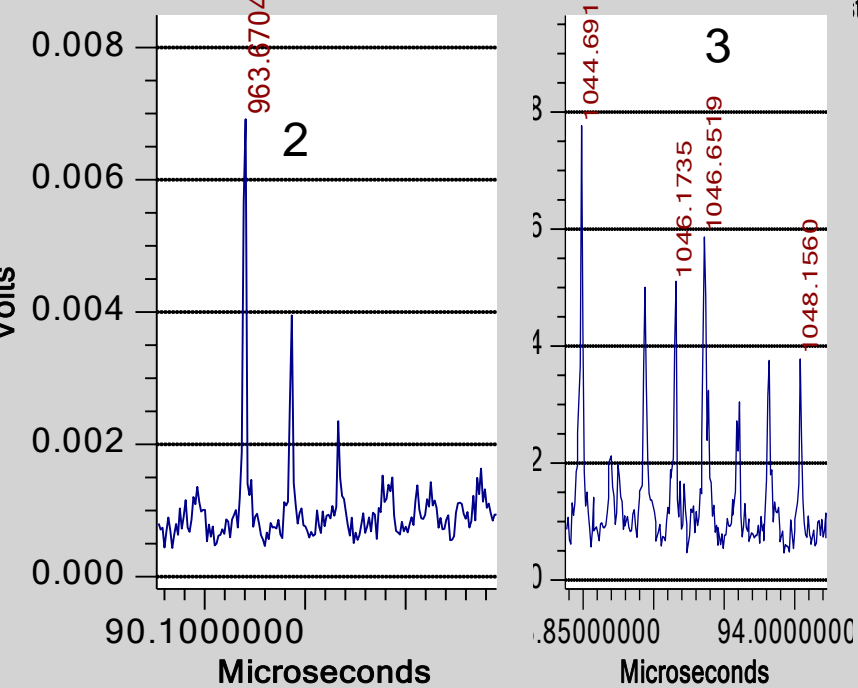
Equimolar mixture diluted in 4-hydroxy- $\alpha$ -cyanocinnamic acid from 1 picomole/ $\mu$ L to 1 attomole/ $\mu$ L

Automated acquisition and calibration averaging 2000 laser shots/spectrum at 1 kHz

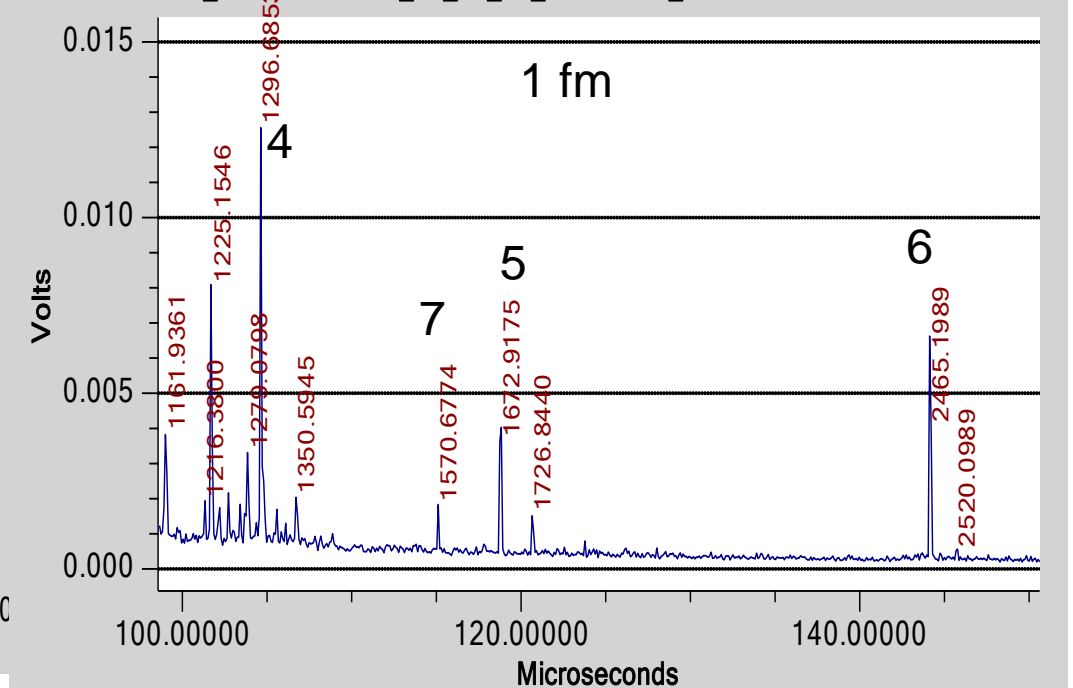
No data processing



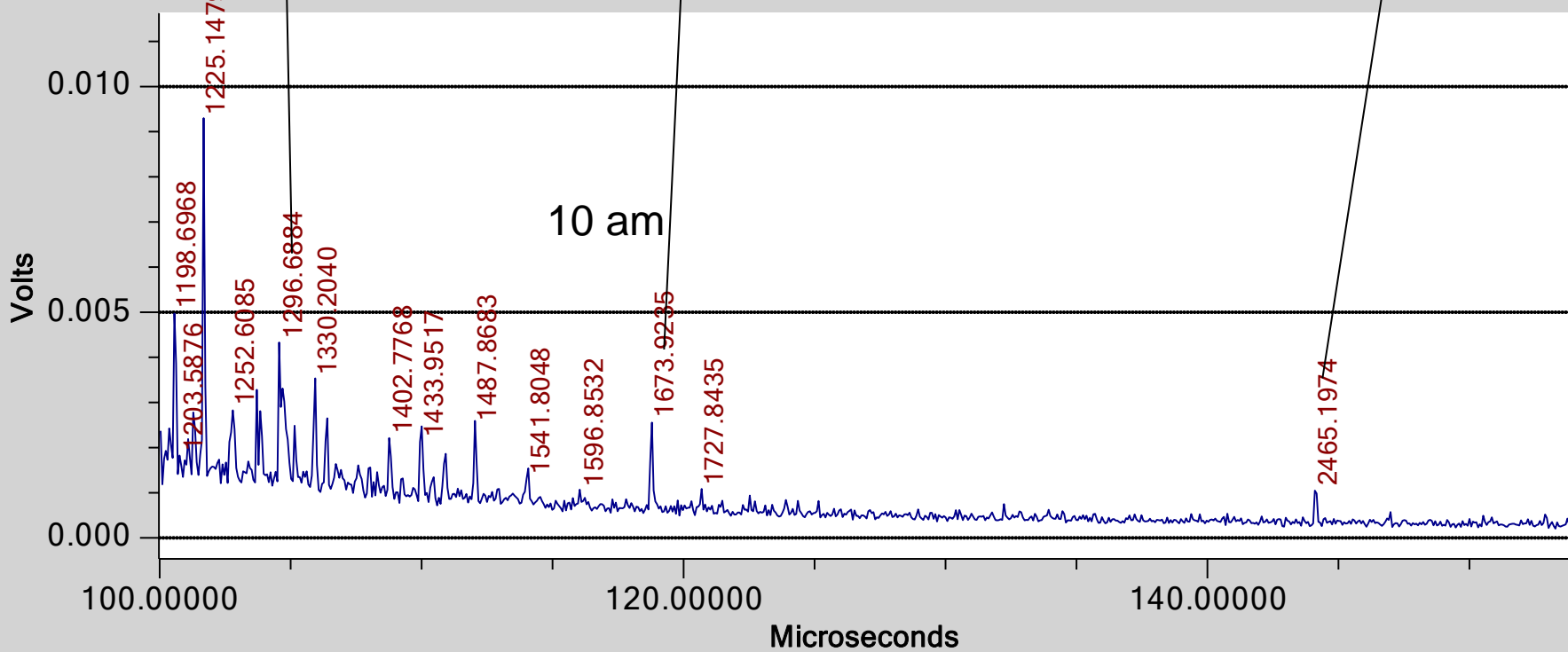
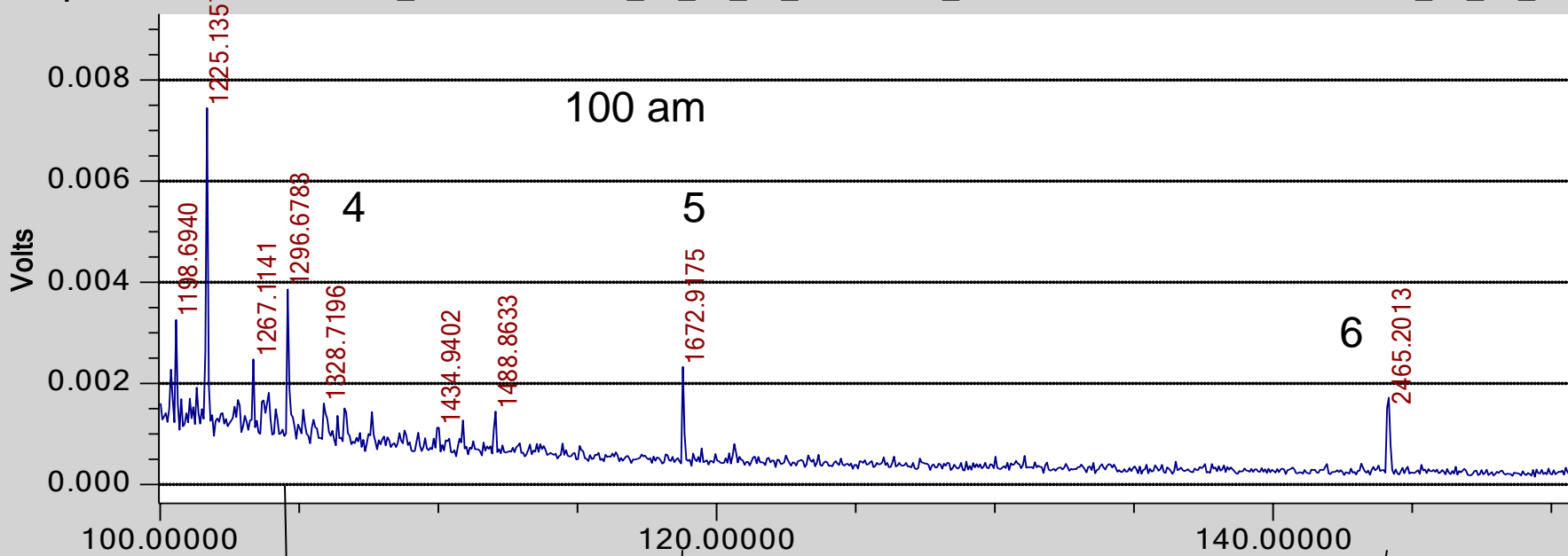
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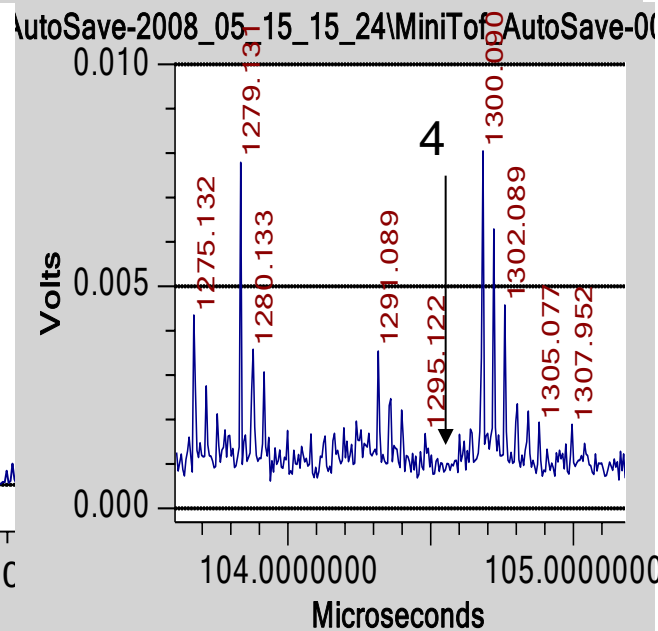
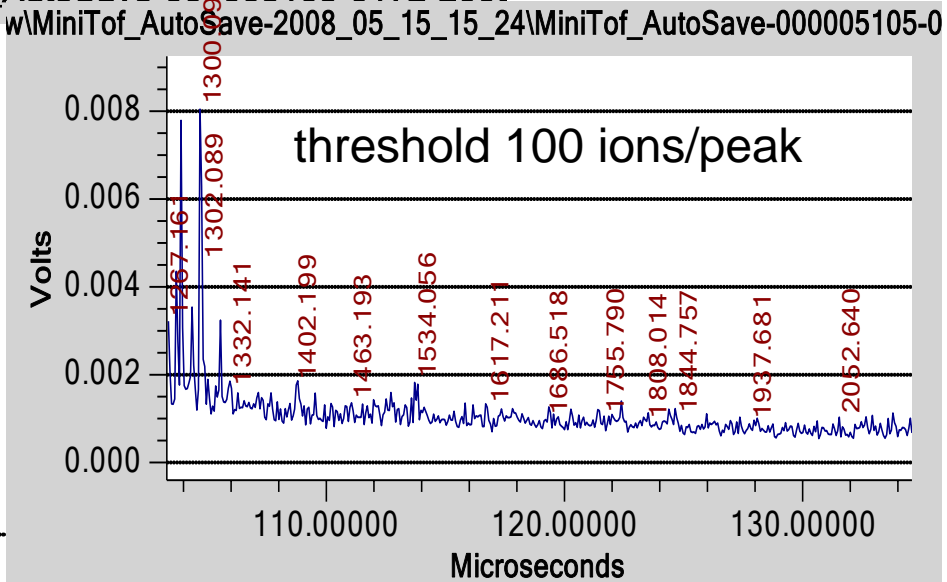
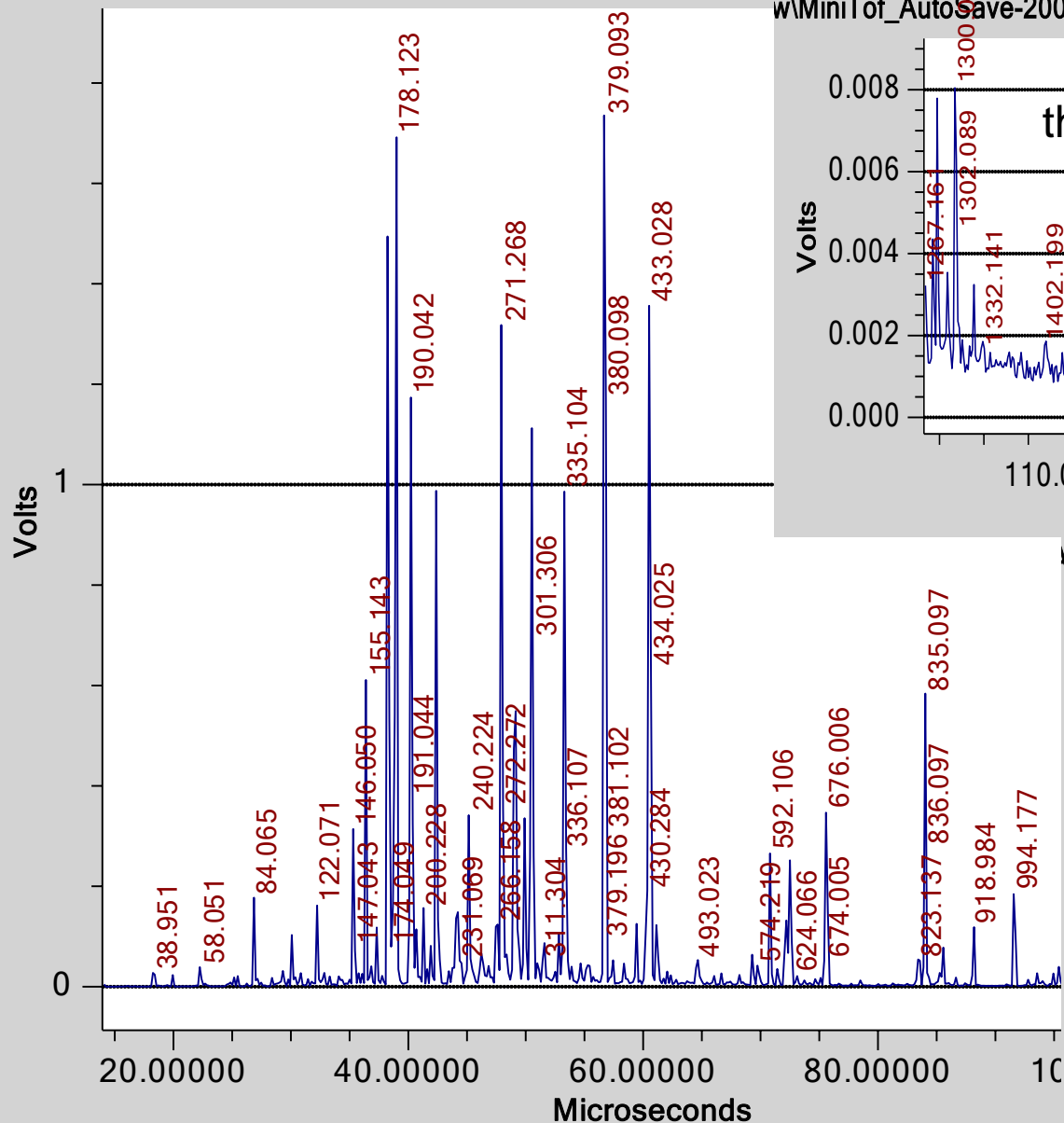


Examples of spectra from serial dilution of peptide standards





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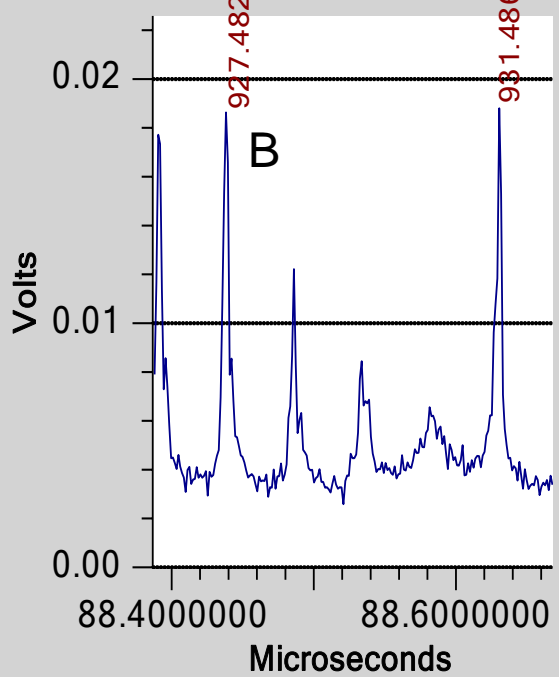
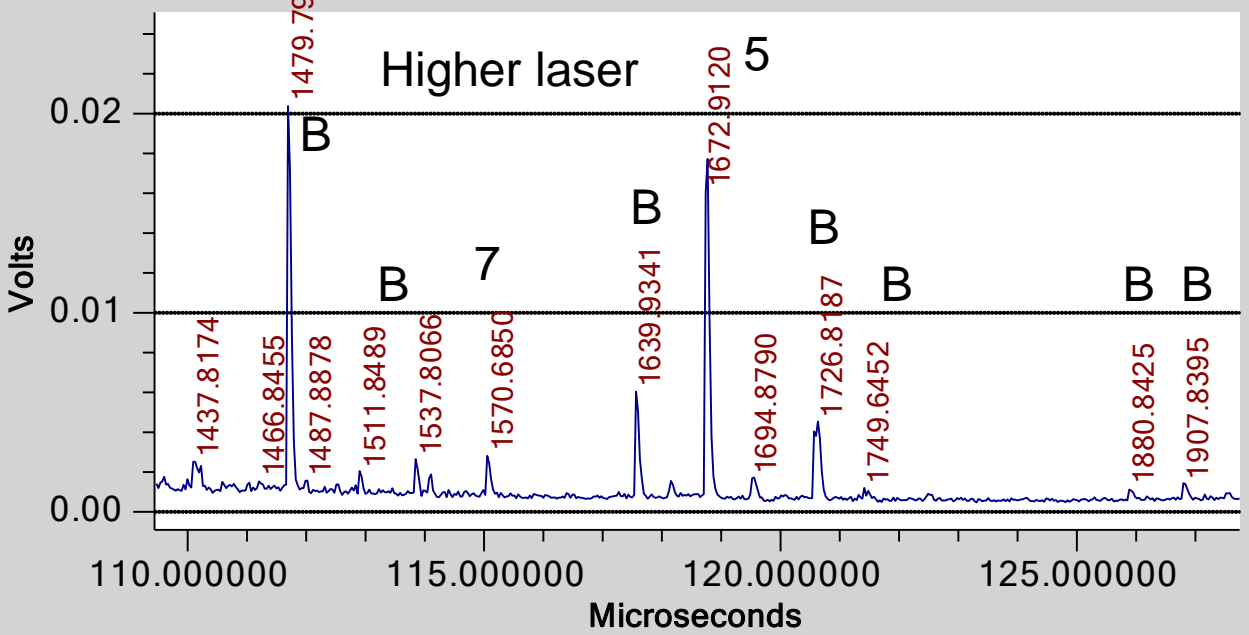
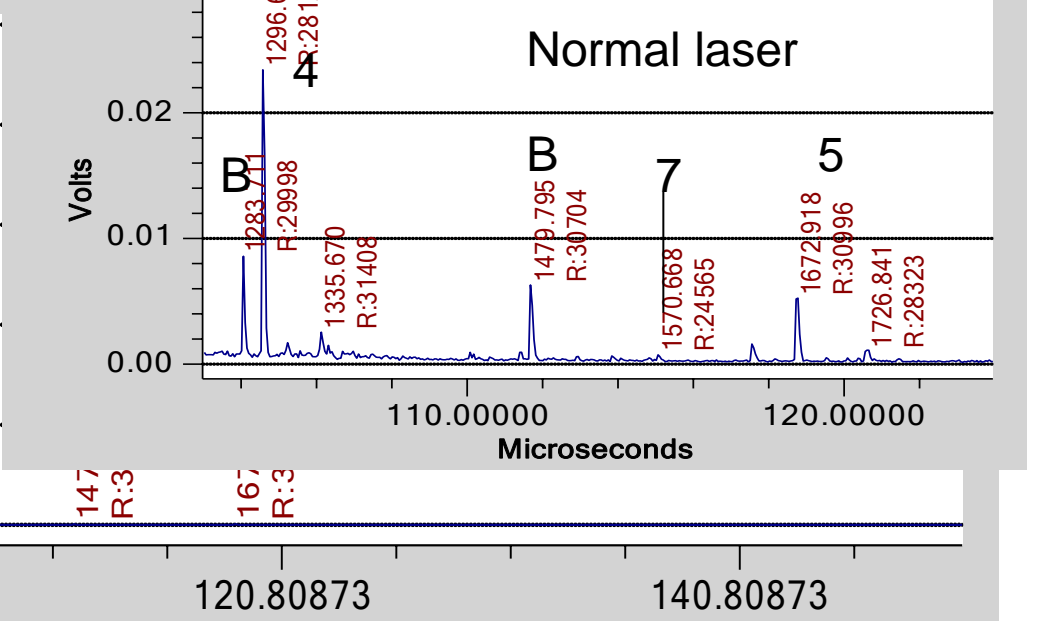
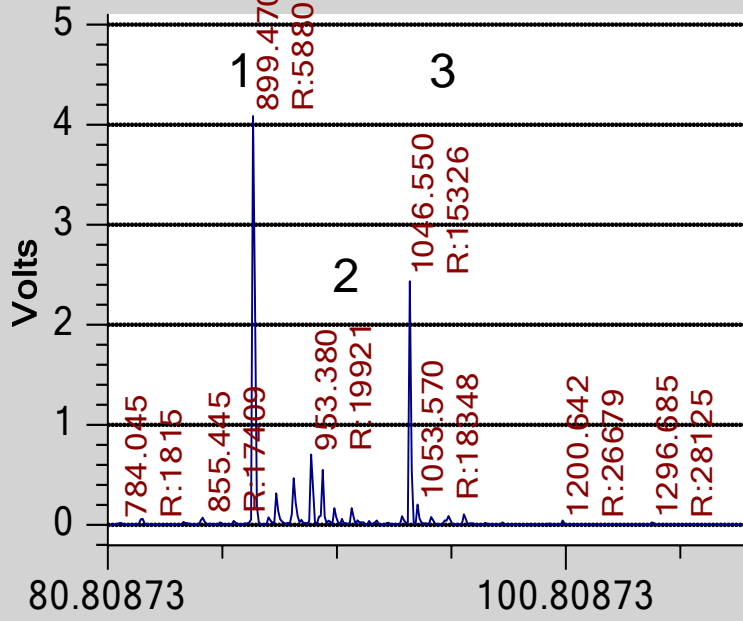


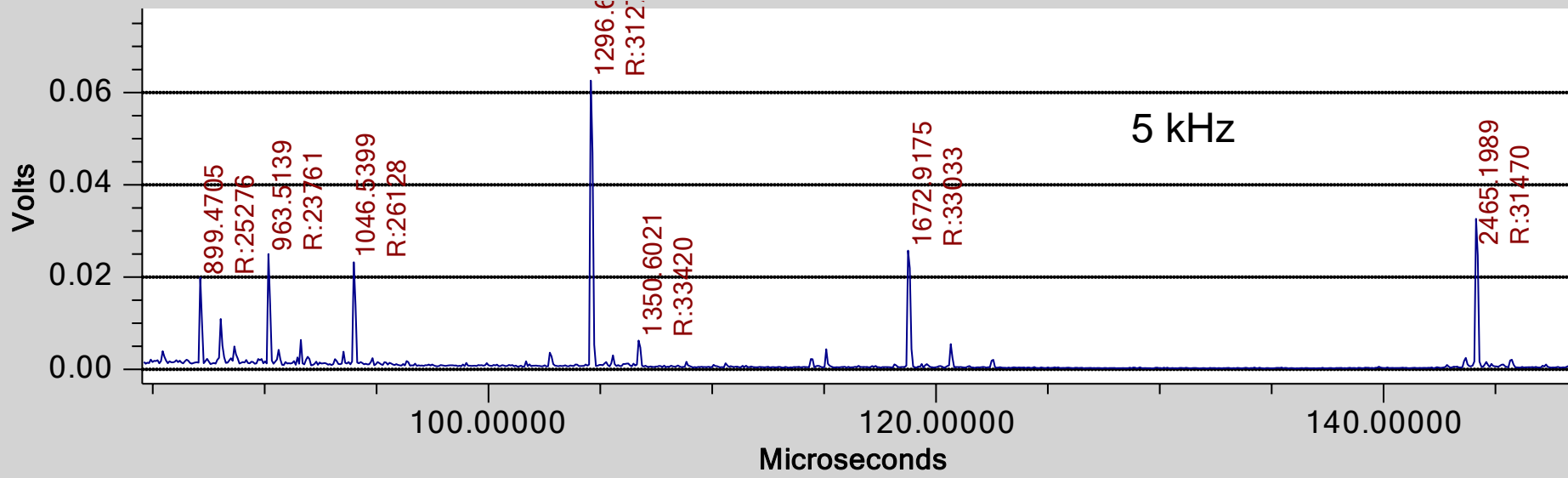
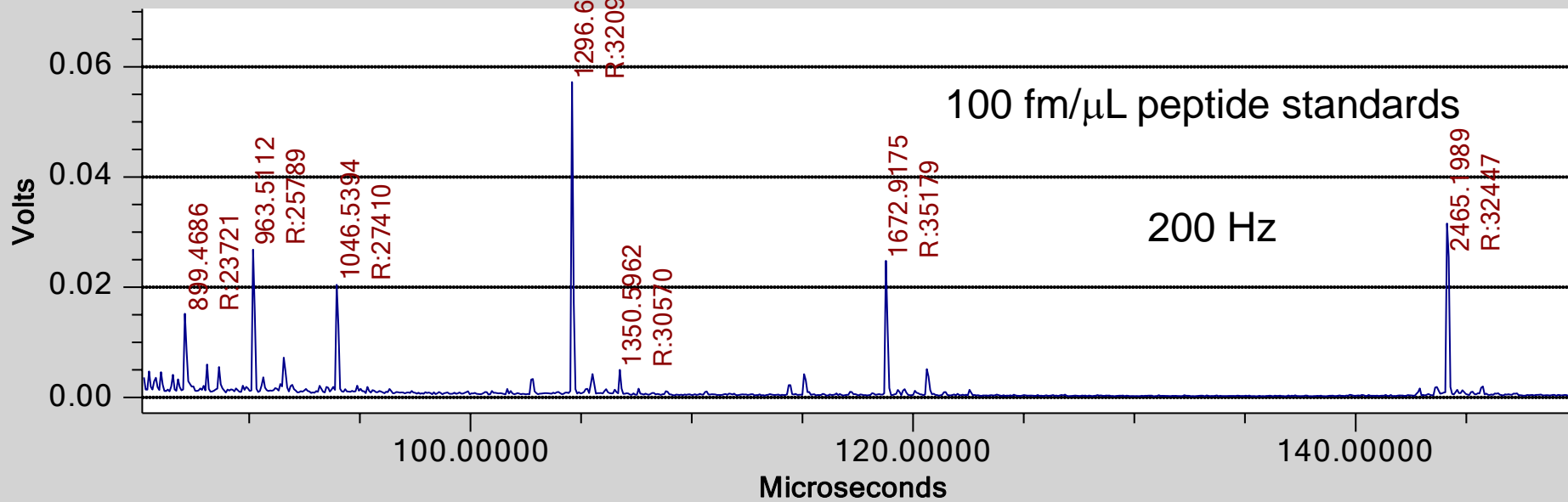
Matrix blank, 2V digitizer range, major matrix peaks off-scale

## Test Mixture for Determining Dynamic Range

<b>Peptide</b>	<b>Conc. (fmole/<math>\mu</math>L)</b>	<b>MH<sup>+</sup></b>	<b>No.</b>
Angiotensin Fragment 1-7	10,000	899.47	1
Angiotensin II Acetate	1,000	1046.54	3
des-Pro <sup>2</sup> Bradykinin	100	963.51	2
Angiotensin I	2	1296.69	4
Neurotensin	1	1672.92	5
Glu-1 Fibrinopeptide B (synthetic)	0.2	1570.67	7
ACTH (18-39)	0.01	2465.20	6
BSA digest	0.1-1	1479.79	B
		927.49	
		1283.71	
		1639.94, etc	







No detectable effect of laser rate on spectrum quality.

# Conclusion and Future

- Present status
  - Resolving power > 30,000 for peptides
  - Mass error < 2 ppm RMS over entire sample plate
  - Detection limit ~1 attomole/ $\mu\text{L}$
  - Dynamic range  $\sim 10^5$
  - Performance independent of laser rate (to 5 kHz)
- Future goals
  - Resolving power >100,000
  - Mass error < 1 ppm RMS
  - 10x improvement in detection limit and dynamic range
  - Isotopic resolution of proteins to 30,000 Da
  - Multiplexed MS-MS with >4000 resolving power for precursor selection and very high sensitivity and throughput
  - Instruments designed for specific applications, e.g. isotope ratios at ppt level

*limited by chemical noise*



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